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Palladium Single Atoms on TiO₂ as a Photocatalytic Sensing Platform for Analyzing the Organophosphorus Pesticide Chlorpyrifos

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Abstract: Traditional methods for analyzing organophosphorus pesticide chlorpyrifos, usually require the tedious sample pretreatment and sophisticated bio-interfaces, leading to the difficulty for real-time analysis. Herein, we use palladium single-atom (PdSA)/TiO₂ as a photocatalytic sensing platform to directly detect chlorpyrifos with high sensitivity and selectivity. PdSA/TiO₂, prepared by an in situ photocatalytic reduction of PdCl₄²⁻ on the TiO₂, shows much higher photocatalytic activity $(10 \text{ molg}^{-1}h^{-1})$ for hydrogen evolution reaction than Pd nanoparticles (1.95 molg⁻¹ h^{-1}), and excellent stability. In the presence of chlorpyrifos, the photocatalytic activity of PdSA/TiO₂ decreases. Through this inhibition effect the platform can realize a detection limit for chlorpyrifos of 0.01 ng mL⁻¹, much lower than the maximum residue limit (10 ppb) permitted by the U.S. Environmental Protection Agency.

Organophosphorus pesticides (OPs) have been historically utilized to eliminate the pests in agricultural industry for high production of crops.^[1] However, the residual pesticides easily enter into human body via the uptake of crops and water as well as the atmosphere. Even a small amount of OPs exposed in vivo can rapidly inhibit the activity of cholinesterase enzymes and subsequently block the neurotransmitter transmission, thus leading to the disfunction of nervous system and the subsequent physical discomfort to people, even death.^[2,3] Chlorpyrifos is one of the representative OPs, and popularly applied to control the agricultural and house hold pests around the world.^[4] The annual consumption of chlorpyrifos in China is up to 18000 tones. The use of chlorpyrifos in such large quantity no doubt lead to the abuse of pesticides. Thus, highly sensitive detection of chlorpyrifos, remained in crops samples and environmental samples, is of vital importance for real-time evaluation of food safety. In terms of chlorpyrifos detection, different analytical methods such as gas/liquid chromatography (GC/LC),^[5,6] GC-MS^[7,8] and nanomaterialsbased biosensors,^[9] have been demonstrated by researchers from various fields. However, the previous strategies still face huge challenges despite the great advances made for the time being. For example, GC/LC and GC-MS usually involve tedious sample pretreatment steps. Biosensing interfaces in nano-biosensors still suffer from bottlenecks of short storage time and harsh storage conditions. Thus these problems hamper the real-time output of experimental results for chlorpyrifos residues to some extent.

Single-atom (SA) catalysts, a newcomer in the field of nanocatalysis, have garnered increasing attention of enormous researchers from various disciplines.^[10] Compared with the conventional catalysts with the size of nanoparticles or nanoclusters, SA catalysts exhibit the atomically dispersed active sites on the support that endows them the optimum atomic utilization efficiency, the excellent electron properties as well as the enhanced catalytic activity.^[11,12] The unique features make SA catalysts be ideal model to understand the catalytic mechanism at the molecule level, simultaneously bridging the gap between homogeneous catalysis and heterogeneous catalysis.^[13] Current researches on SA catalysts are mainly centered on the energy-related field, such as electro-

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Communications

catalyst and photocatalyst.^[14,15] Previous reports have fully demonstrated the extraordinary activity and stability of SA catalysts in hydrogen evolution reaction (HER),^[16,17] oxygen reduction reaction (ORR),^[18] and oxygen evolution reaction (OER).^[19] Despite booming growth regarding energy-related application, the bioapplication of SA catalysts is still in its infancy,^[20,21] and SA catalyst applied for detection of pesticides residues was never reported until now.

Herein, we first adopt PdSA supported TiO_2 as an advanced photocatalytic sensing platform for highly sensitive detection of chlorpyrifos via restricting the photocatalytic H₂ evolution activity of PdSA by chlorpyrifos. Benefiting from the highly uniform and atomic dispersion of Pd active sites, PdSA/TiO₂ exhibits the much enhanced photocatalytic HER performance and adequate sensitivity for chlorpyrifos determination. Without any complex pre-processing procedures, such a method is less time-consuming and direct, providing great potential for environmental monitoring chlorpyrifos residues.

The SA catalyst of Pd/TiO_2 was prepared by an in situ photocatalytic reduction strategy under the iced state. The homogeneous mixture solution of TiO_2 and Pd precursors (K_2PdCl_4) was treated with

liquid nitrogen to form an ice bulk. The strong interaction between the electron-rich oxygen of TiO_2 and the empty orbitals of the metal Pd, deposited the PdCl_4^{2-} on the surface of TiO_2 nanoparticles. Irradiation with Xe light of the ice bulk, excited TiO_2 to produce photogenerated electrons that subsequently promoted the reduction of PdCl_4^{2-} to form metal Pd species, giving Pd_1/TiO_2 . As a control, the same mixture, but without the freezing process was directly irradiated by Xe light, to give Pd_2/TiO_2 .

Transmission electron microscope (TEM), scanning transmission electron microscope (STEM)-energy dispersive Xray spectroscopy (EDS) and the aberration-corrected highangle annular dark-field (HAADF)-STEM were applied to characterize the morphology of the as-made Pd species. TEM images demonstrate that there are many small dots on the surface of Xe light-irradiated TiO2, Pd1/TiO2 and Pd2/TiO2 (Figure S1 in the Supporting Information) when compared with the untreated TiO₂. The results from STEM-EDS and the HAADF-STEM characterizations clearly show that metal Pd in Pd_1/TiO_2 sample is uniformly dispersed onto TiO_2 substrate (Figure 1 a,b). The X-ray absorption near-edge structure (XANES) spectroscopy of Pd₁/TiO₂ with atomic dispersion show a slightly higher energy for the absorption edge than that of Pd foil (Figure 1d), suggesting the presence of oxidized Pd. Furthermore, the Pd K-edge spectra in extended X-ray adsorption fine structure (EXAFS) analysis emerge a sharp characteristic peak of Pd-O bond (in the region of 1 Å to 2 Å) and a weak peak of Pd–Pd (Figure 1 c), fully confirming the subjective status of PdSA on TiO₂ substrate after the iced treatment. In X-ray diffraction (XRD) characterization, Pd₁/TiO₂ does not exhibit any peak of Pd nanoparticles when compared with Pd₂/TiO₂ sample (Figure S2), further demonstrating that PdSA with the ultra-



Figure 1. The structural characterizations of $PdSA/TiO_2$. a) EDS-STEM elemental mapping and b) HAADF-STEM image of $PdSA/TiO_2$, c) Fourier transform of the Pd K-edge of the EXAFS spectra of $PdSA/TiO_2$, d) normalized XANES spectra at the Pd L3-edge of $PdSA/TiO_2$ and Pd foil.

small size has no significant effect on the structure of TiO₂. Thus, Pd_1/TiO_2 is called PdSA/TiO₂ while Pd_2/TiO_2 is named for Pd nanoparticles/TiO₂ (PdNPs/TiO₂). The Pd contents in PdSA/TiO₂ and PdNPs/TiO₂ were determined to be 0.53 wt % and 0.56 wt %, respectively via the inductively coupled plasma-atomic emission spectroscopy (ICP-AES).

Figure 2 a shows the schematic principle on photocatalytic HER of PdSA/TiO₂. Under the irradiation of visible light, the electrons from valence band of TiO_2 was excited into the



Figure 2. a) The principle of the PdSA/TiO₂-based sensing platform utilizing photocatalytic H₂ production; b) The photocatalytic H₂ production rate of PdSA/TiO₂ and PdNPs/TiO₂; c) The inhibition behavior of chlorpyrifos on the photocatalytic HER performance of both PdSA/TiO₂ and PdNPs/TiO₂.

conduction band that directly reacted H⁺ in the solution to generate H₂ through PdSA. Meanwhile, the generated positive charged hole (h⁺) promoted the oxidation of ethanol to form acetaldehyde. Figure 2b displays the photocatalytic performance of PdSA/TiO₂ and PdNPs/TiO₂. As expected, the TiO₂ support substrate, did not present any photocatalytic HER signal due to the lack of metal Pd. PdSA/TiO₂ exhibits the remarkably enhanced photocatalytic performance with the H₂ production rate of up to $10 \text{ mol g}^{-1}\text{h}^{-1}$, much higher than that of PdNPs/TiO₂ (1.95 mol $g^{-1}h^{-1}$). Such an excellent photocatalytic performance generated at a lower loading level of Pd further suggested the super-high density and active sites of PdSA on TiO₂ substrate in comparison to PdNPs. Most importantly, we unexpectedly discover that in the presence of chlorpyrifos, the photocatalytic HER activity of PdSA/TiO₂ is significantly inhibited whereas that of PdNPs/ TiO_2 did not exhibit any observed change (Figure 2a,c). Though both PdSA/TiO₂ and PdNPs/TiO₂ have the similar loading amount of Pd element, PdSA exhibited much higher dispersion of Pd active sites due to the ultimately atomic utilization efficiency than PdNPs. Chlorpyrifos is prone to block the photocatalytic HER active sites of PdSA, thus suggesting that PdSA/TiO₂ is more sensitive toward chlorpyrifos.

X-ray photoelectron spectra (XPS) and mass spectra (MS) were employed to characterize the interaction between chlorpyrifos and PdSA/TiO₂. Figure 3 displays the XPS spectra of Pd 3d. For the as-synthesized PdSA/TiO₂, the binding energies of Pd0 3d at 341.28 $(3d_{\scriptscriptstyle 3/2})~eV$ and 336.08 eV $(3d_{5/2})$ exhibit slight difference from the standard values of bulk Pd $(3d_{3/2} = 341.1 \text{ eV}; 3d_{5/2} = 335.2 \text{ eV})$ because of the interaction between Pd species and TiO2. In the meantime, the peaks in XPS spectrum of Pd 3d at 342.48 eV and 337.18 eV typically verify the existence of oxidized Pd^{II} (Figure 3a), being in good agreement with the XANES result. In XPS analysis of the PdSA/TiO2 exposed to chlorpyrifos, both the peaks of Pd $3d_{3/2}$ and Pd $3d_{5/2}$ show noticeable negative shift compared with that of the unexposed PdSA/TiO₂ owing to the adsorption of chlorpyrifos with electron-rich group on the electron-withdrawing PdSA/ TiO₂ (Figure 3b). This typical XPS feature of Pd 3d before and after chlorpyrifos exposure is highly in accordance with the previous report on the strong interaction of Pd^{II} with sulfur-containing compound or nanomaterials.^[22,23] In MS



Figure 3. XPS of a) Pd 3d from PdSA/TiO₂ and b) Pd 3d XPS of PdSA/TiO₂ before and after the reaction with chlorpyrifos.

analysis, $Pd(CH_3CN)_4(BF_4)_2$ was used here as Pd^{II} species to react with chlorpyrifos. One peak with 1⁺ charge state is observed in electrospray ionization mass spectrometry (ESI-MS), due to loss of the counterion, BF_4^- . After deconvolution of m/z, the average measured composition of S1 is 977.76 Da, which exactly agreeing with the molecular composition of S1 $[(C_9H_{11}Cl_3NO_3PS)_2Pd(BF_4)_2]$. The corresponding fragmentation masses are around 890.76 (Figure S3), indirectly confirming the strong interaction of chlorpyrifos with PdSA on TiO₂ supports.

The developed PdSA/TiO₂-based photocatalytic sensing platform provides a direct and simple strategy for chlorpyrifos determination. Some certain parameters were explored subsequently to evaluate the reliability of this method. As shown in Figure S4, the photocatalytic H₂ production rate of PdSA/ TiO₂ rapidly increases in the first 10 min of Xe light irradiation and reaches a plateau at 30 min. Hence, 30 min is the optimal time selected to perform this photocatalytic reaction. Besides, the reaction time of 30 min implies that this approach is adequate to realize the real-time detection of chlorpyrifos residues. The photocatalytic stability of PdSA/ TiO₂ was also investigated in detail. The synthesized PdSA/ TiO₂ was stored in the centrifuged tube at room temperature, and tested every two months. It is demonstrated that the photocatalytic H₂ production rate of PdSA/TiO₂ retains the initiate level well even after eight months (Figure S5). Especially, the same dose of chlorpyrifos still realizes the same inhibition effect on the photocatalytic performance of PdSA/TiO₂ (Figure 4a). These results underscore the excellent stability of PdSA/TiO2-based photocatalytic sensor as well as the favorable prospect toward chlorpyrifos analysis. Inspired by the above results, this PdSA/TiO₂-based photocatalytic sensing strategy was then applied to detect different concentrations of chlorpyrifos standard samples. The resulting photocatalytic H₂ production rate decrease with increasing chlorpyrifos content in the samples from 0.03 ngmL⁻¹ to $10 \,\mu g \,m L^{-1}$ (Figure 4b). The detection of limit is determined to be lower to 0.01 ngmL^{-1} , much lower than those of the previously reported methods.^[24,25] Moreover, according to the report from U.S. Environment Protection Agency, the maximum residues amount of chlorpyrifos in nearly all the agriculture products has to be lower than 10 ppb.^[26] Thus, our proposed photocatalytic sensing platform features the remarkable sensitivity for chlorpyrifos determination.

> Three other OPs commonly used in crop protection, including paraoxon-ethyl, monocrotophos, and dipterex were examined to evaluate the selectivity of this method. The results showed that these other pesticides did not cause any obvious inhibition effect on the photocatalytic activity of PdSA/TiO₂ (Figure 4c). Moreover, some anions and cations that are ubiquitous on vegetables and crops have been successfully verified to have no influence on the photocatalytic performance of PdSA/TiO₂ (Figure 4d), significantly certifying the reliability of the proposed method for chlorpyrifos assay. Spinach was used here as the model vegetable to explore the practical use of this sensing platform. The results achieved demonstrate that the vegetable samples



Figure 4. a) The photocatalytic HER stability of PdSA/TiO₂ before and after the introduction of chlorpyrifos. b) The typically photocatalytic response of PdSA/TiO₂ toward different concentrations of chlorpyrifos, 0.03 ng mL⁻¹, 1 ng mL⁻¹, 30 ng mL⁻¹, 200 ng mL⁻¹, 1 µg mL⁻¹, 10 µg mL⁻¹. c) The interference assessment of other commonly used organophosphorus pesticides on chlorpyrifos determination. d) The interference analysis of the anions and cations common in vegetable samples.

before and after spiking with chlorpyrifos exhibit distinct inhibition behaviors to the photocatalytic activity of PdSA/TiO₂, with the average recovery rate of 125% (Figure S6 and Table 1), exhibiting the acceptable practicability for real-time assay of chlorpyrifos residues.

 $\label{eq:table_transform} \begin{array}{l} \textbf{Table 1:} \\ \text{Recovery measurement of chlorpyrifos-spiked vegetable samples by PdSA/TiO_2-based photocatalytic sensing platform.^{[a]} \end{array}$

Property		Sample number				
	1	2	3	4	5	
Chlorpyrifos spiked (ng/mL)	0	5	50	200	1000	
Chlorpyrifos detected (ng/mL)	0.03	6.9	62	238	1200	
Recovery (%)	-	138	124	119	120	

[a] (ng/mL) indicates ng of chlorpyrifos to mL of 25 % ethanol aqueous solution.

In conclusion, we developed an advanced photocatalytic sensing platform that employed SA catalysts of PdSA/TiO₂ for direct and simple chlorpyrifos detection. HAADF-STEM and XANES characterizations fully confirmed the monoatomic feature of as-made PdSA/TiO₂. PdSA/TiO₂, owing to their high number of active sites, displayed much higher photocatalytic HER performance than PdNPs/TiO2. Most especially, the presence of chlorpyrifos has a more pronounced inhibition effect on the photocatalytic activity of PdSA/TiO₂ in comparison to PdNPs/TiO₂. The PdSA/TiO₂based photocatalytic sensing platform achieves a good linear relationship ranged from 0.03 ngmL^{-1} to $10 \mu \text{gmL}^{-1}$ chlorpyrifos with a very low detection limit of 0.01 ng mL^{-1} . Moreover, both the commonly existing ions and other generally used OPs (paraoxon, monocrotophos and dipterex) did not interfere with chlorpyrifos determination, indicative of the good selectivity of PdNPs/TiO₂-based sensing of chlorpyrifos. For chlorpyrifos-spiked spinach samples, the proposed sensing method exhibited definite feasibility with the average recovery rate of 125%. This typical SA catalyst-based photocatalytic sensing platform opens up a new pathway for chlorpyrifos residual detection, providing the prospect for exploring novel biosensing strategies for OPs.

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Conflict of interest

The authors declare no conflict of interest.

Keywords: chlorpyrifos detection · hydrogen evolution reaction · photocatalysis · single-atom catalyst · titanium

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- [1] D. K. Nomura, J. K. Casida, J. Agric. Food Chem. 2011, 59, 2808–2815.
- [2] D. Lu, J. Wang, L. Wang, D. Du, C. Timchalk, R. Barry, Y. Lin, *Adv. Funct. Mater.* 2011, 21, 4371–4378.
- [3] X. Ge, W. Zhang, Y. Lin, D. Du, Biosens. Bioelectron. 2013, 50, 486–491.
- [4] L. Wang, D. Lu, J. Wang, D. Du, Z. Zou, H. Wang, J. N. Smith, C. Timchalk, F. Liu, Y. Lin, *Biosens. Bioelectron.* 2011, 26, 2835– 2840.
- [5] A. Shanker, C. Sood, V. Kumar, S. D. Ravindranath, Pest Manage. Sci. 2001, 57, 458-462.
- [6] S. Samadia, H. Sereshtia, Y. Assadi, J. Chromatogr. A 2012, 1219, 61-65.
- [7] K. Maštovská, S. J. Lehotay, J. Chromatogr. A 2004, 1040, 259– 272.
- [8] H. Kataoka, H. L. Lord, J. Pawliszyn, J. Chromatogr. A 2000, 880, 35-62.
- [9] W. Zhang, A. M. Asiri, D. Liu, D. Du, Y. Lin, *TrAC Trends Anal. Chem.* 2014, 54, 1–10.
- [10] B. Qiao, A. Wang, X. Yang, L. F. Allard, Z. Jiang, Y. Cui, J. Liu, J. Li, T. Zhang, *Nat. Chem.* **2011**, *3*, 634.
- [11] J. Qian, Z. Yang, C. Wang, K. Wang, Q. Liu, D. Jiang, Y. Yan, K. Wang, J. Mater. Chem. A 2015, 3, 13671–13678.
- [12] J. Chen, Y. Huang, P. Kannan, L. Zhang, Z. Lin, J. Zhang, T. Chen, L. Guo, Anal. Chem. 2016, 88, 2149–2155.
- [13] W. Hu, Q. Chen, H. Li, Q. Ouyang, J. Zhao, *Biosens. Bioelectron.* 2016, 80, 398–404.
- [14] Y. Chen, S. Ji, Y. Wang, J. Dong, W. Chen, Z. Li, R. Shen, L. Zheng, Z. Zhuang, D. Wang, Y. Li, *Angew. Chem. Int. Ed.* 2017, 56, 6937–6941; *Angew. Chem.* 2017, 129, 7041–7045.
- [15] W. Zhong, R. Sa, L. Li, Y. He, L. Li, J. Bi, Z. Zhuang, Y. Yu, Z. Zou, J. Am. Chem. Soc. 2019, 141, 7615-7621.





- [16] K. Jiang, B. Liu, M. Luo, S. Ning, M. Peng, Y. Zhao, Y. Lu, T. Chan, F. Groot, Y. Tan, *Nat. Commun.* 2019, 10, 1743.
- [17] P. Zhou, F. Lv, N. Li, Y. Zhang, Z. Mu, Y. Tang, J. Lai, Y. Chao, M. Luo, F. Lin, J. Zhou, D. Su, S. Guo, *Nano Energy* **2019**, *56*, 127–137.
- [18] S. Yang, J. Kim, Y. J. Tak, A. Soon, H. Lee, Angew. Chem. Int. Ed. 2016, 55, 2058–2062; Angew. Chem. 2016, 128, 2098–2102.
- [19] J. Wang, Z. Li, Y. Wu, Y. Li, Adv. Mater. 2018, 30, 1801649.
- [20] L. Wang, J. Chen, L. Gan, J. Wang, S. Dong, Sci. Adv. 2019, 5, eaav5490.
- [21] L. Jiao, H. Yan, Y. Wu, W. Gu, C. Zhu, D. Du, Y. Lin, Angew. Chem. Int. Ed. 2019, https://doi.org/10.1002/anie.201905645; Angew. Chem. 2019, https://doi.org/10.1002/ange.201905645.
- [22] G. Celik, S. Ailawar, S. Gunduz, J. Miller, P. Edmiston, U. Ozkan, *Ind. Eng. Chem. Res.* 2019, 58, 4054–4064.
- [23] Z. Luo, Y. Ouyang, H. Zhang, M. Xiao, J. Ge, Z. Jiang, J. Wang, D. Tang, X. Cao, C. Liu, W. Xing, *Nat. Commun.* **2018**, *9*, 2120.
- [24] Q. Liu, Y. Yin, N. Hao, J. Qian, L. Li, T. You, H. Mao, K. Wang, Sens. Actuators B 2018, 260, 1034–1042.
- [25] H. Li, J. Li, Q. Xu, X. Hu, Anal. Chem. 2011, 83, 9681-9686.
- [26] K. Zhang, Q. Mei, G. Guan, B. Liu, S. Wang, Z. Zhang, Anal. Chem. 2010, 82, 9579–9586.

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